

Pulmonary Toxicity of Intratracheally Instilled Multiwall Carbon Nanotubes in Male Fischer 344 Rats

Shigetoshi AISO^{1*}, Kazunori YAMAZAKI¹, Yumi UMEDA¹, Masumi ASAKURA¹,
Tatsuya KASAI¹, Mitsutoshi TAKAYA², Tadao TOYA², Shigeki KODA²,
Kasuke NAGANO¹, Heihachiro ARITO¹ and Shoji FUKUSHIMA¹

¹Japan Bioassay Research Center, Japan Industrial Safety and Health Association, 2445 Hirasawa, Hadano, Kanagawa 257-0015, Japan

²National Institute of Occupational Safety and Health, 6–21–1 Nagao, Tama-ku, Kawasaki 214-8585, Japan

Received July 23, 2009 and accepted January 8, 2010

Abstract: In order to assess pulmonary toxicity of multiwall carbon nanotubes (MWCNT), male F344 rats were intratracheally instilled with MWCNT suspension at a dose of 40 or 160 $\mu\text{g}/\text{head}$ or α -quartz particles as a positive control at a dose of 160 $\mu\text{g}/\text{head}$ and sacrificed for lung histopathology and bronchoalveolar lavage (BAL) fluid analyses on Day 1, 7, 28 or 91 after instillation. Well-dispersed MWCNT brought about dose- or time-dependent changes in lung weight, total proteins, albumin, lactate dehydrogenase and alkaline phosphatase in the BAL fluid, and pulmonary lesions including inflammation, Type II cell hyperplasia, microgranulomas and fibrosis. Phagocytosed and free forms of MWCNT were found in both bronchiolar and alveolar spaces. MWCNT deposition in the bronchus-associated lymphoid tissue gradually increased after instillation. Persistent infiltration of macrophages, transient infiltration of inflammatory cells primarily composed of neutrophils, microgranulomas associated with macrophages engulfing MWCNT, Type II cell hyperplasia and fibrosis with alveolar wall thickening as well as number of multinucleated alveolar macrophages increased dose-dependently. The MWCNT-induced lesions were more potent on Day 91 than the α -quartz-induced ones at an equal mass dose. The present results for intratracheally instilled MWCNT were extrapolated to potential inhalation exposure of humans to MWCNT at workplaces based on several assumptions.

Key words: Multiwall carbon nanotube, Neutrophils, Alveolar macrophages, Granuloma, Fibrosis, Alveolar wall thickening, Type II cell hyperplasia, Multinucleated cell

Introduction

Carbon nanotubes (CNT) have been reported to possess unusually excellent electrical, mechanical and thermal properties, and thus have many potential applications in electronics, computers, aerospace, and the medical and pharmaceutical industries^{1, 2}. According to an article in *Nature*³, Smalley predicted that in time, millions of tons of CNT will be produced worldwide every year. Yearly production volumes of single-wall

carbon nanotubes (SWCNT) and multiwall carbon nanotubes (MWCNT) in Japan were estimated to be 0.1 and 60 tons, respectively⁴. With the rapid increase in industrial production of CNT, much concern has been raised over the health consequences for workers who are exposed to SWCNT or MWCNT in their occupational settings. Neither epidemiological nor medical case studies have been reported on the health consequences of CNT-exposed workers. However, the toxicity of CNT has been examined by exposing experimental animals to CNT by intraperitoneal injection, intratracheal instillation, pharyngeal aspiration or by inhalation. The reported findings of these *in vivo* toxicity studies of CNT

*To whom correspondence should be addressed.
E-mail: s-aiso@jisha.or.jp

included mesotheliomas in *p53* gene-deficient mice⁵) and intact male F344 rats⁶), asbestos-like pathogenicity in female mice⁷), induction of inflammation and fibrosis of the lung in mice^{8–11}) and rats^{12, 13}), inflammation and oxidative stress in the lungs of vitamin E-deficient mice¹⁴), granulomas in the lungs of rats and mice^{5–15}), induction of apoptosis in the absence of inflammation in the rat lung¹⁶), and systemic immunosuppression in mice¹⁷). Inflammation was reported to occur in the mouse lung after intratracheal instillation^{8–11, 14}) but not after inhalation exposure to MWCNT¹⁷). The pulmonary toxicity of MWCNT was reported to alter with pretreatment of MWCNT¹⁸). Mutation of *k-ras* gene locus in the lungs of mice exposed by inhalation to SWCNT¹¹), and positive clastogenicity and aneugenicity of MWCNT¹⁹) in Type II pneumocytes of female rats given intratracheally instilled MWCNT have also been reported. In addition to positive mutagenicity, oxidative stress and pulmonary lesions such as inflammation, hyperplasia and fibrosis are important determinants in carcinogenesis^{20, 21}).

The present study was designed to examine dose- and time-dependent relationships between deposition of MWCNT and its extent in the lung, and pulmonary toxic responses such as inflammation, granuloma and fibrosis in rats intratracheally instilled with MWCNT. The affected lungs were examined for both lung histopathology and biochemical and cytological analyses of bronchoalveolar lavage (BAL) fluid, with reference to the lung lesions and their severities induced by intratracheally instilled α -quartz as a positive control. The results of the dose characteristics of MWCNT in the suspension were accepted for publication in this Journal as a separate paper²²).

Materials and Methods

Animals

Male F344/DuCrI CrIj rats were purchased from Charles River Japan, Inc. (Kanagawa, Japan) at the age of 11 wk. The animals were quarantined and acclimated for 2 wk, then housed individually in stainless steel wire-mesh hanging cages (170W × 294D × 176H mm) under controlled environmental conditions (temperature of 24 ± 2°C and a relative humidity of 55 ± 10% with 15 to 17 air changes/h). Fluorescent lighting was controlled automatically to provide a 12-h light/dark cycle. All rats had free access to sterilized water and γ -irradiation-sterilized commercial pellet diet (CRF-1, Oriental Yeast Co., Ltd., Tokyo, Japan). The animals were cared for in accordance with the Guide for the Care and Use of Laboratory Animals²³), and the present study was approved by the ethics committee of the

Japan Bioassay Research Center (JBRC).

Test substances

MWCNT was kindly supplied by MITSUI & Co. Ltd. (MWCNT-7, Lot No. 061220, Tokyo, Japan). α -Quartz in the form of crystalline silica (MIN-U-SIL 5) was purchased from US. Silica Co. (WV, USA), and the nominal size of α -quartz particles was 1.7 μ m in median diameter. MWCNT and α -quartz were used in the present study as produced; i.e., without being purified or further sieved. Since neither MWCNT nor α -quartz is water-soluble or wettable, these test substances were suspended in phosphate-buffered saline (PBS) containing 0.1% Tween 80 as a colloidal dispersant and subjected to ultrasonication for 20 min with an ultrasonic homogenizer (VP-30S, 20 kHz, 300 W, TAITEC Co., Ltd, Tokyo, Japan). The size distribution of the MWCNT in the suspension was measured by both a dynamic light scattering size measurement (DLS) and scanning electron microscopic (SEM) observation²²). Briefly, the DLS measurement showed that the median hydrodynamic diameter of MWCNT in the suspension after 20-min ultrasonication ranged below 1.0 μ m. The SEM observation revealed that the mean length and width of the MWCNT fibers were 5.0 μ m and 88 nm, respectively, and that fibers longer than 5.0 μ m occupied 38.9% of the total fibers counted. The MWCNT was found to contain 4,400 ppm (wt/wt) iron, 48 ppm chromium and 17 ppm nickel by graphite furnace atomic absorption spectrometric analysis, and these levels of metals were considered not to elicit any positive pulmonary responses²²). Measurement of endotoxin levels in the PBS-Tween 80 suspending MWCNT or α -quartz was commissioned to Japan SLC, Ltd. (Tokyo, Japan). The levels of endotoxin in the PBS-Tween 80 suspending MWCNT, α -quartz and in the vehicle PBS-Tween 80 were below 0.9 pg/ml, indicating the absence of contamination of MWCNT or α -quartz with bacteria.

Intratracheal instillation

Immediately before intratracheal instillation, the ultrasonicated suspension of MWCNT, α -quartz or vehicle solution was further subjected to additional ultrasonication for 30s with a sonicator (US-2, AS ONE Co., Ltd., Tokyo, Japan). After inhalational anesthetization with isoflurane gas (Forane, Abbott Japan Co., Ltd., Tokyo, Japan), the suspension of MWCNT or α -quartz in PBS-Tween 80 (0.3 ml) or the vehicle solution (0.3 ml) was intratracheally instilled, using a microsyringe cannula of Intratracheal Aerosolizer (1A-1B, PennCentury, Inc., USA). Successful delivery of the instilled MWCNT into lungs was confirmed by observing both a wheezing sound and rapid recovery from anesthesia with nei-

ther abnormal behavior nor negative health outcomes at the time of instillation. The animals treated with the MWCNT, α -quartz suspension or the vehicle solution recovered quickly after anesthesia without any behavioral or negative health outcomes.

Experimental design

A total of 192 rats were used for the study of MWCNT toxicity, while 96 rats were used for the positive control study of α -quartz toxicity, consisting of 48 α -quartz-dosed rats and 48 vehicle-dosed rats for a negative control. After quarantine and acclimation for 2 wk, the animals were divided by stratified randomization into two MWCNT-dosed, one α -quartz-dosed positive and one vehicle-dosed negative control group, each consisting of 16 rats/group, at each of the time points for sacrifice. The two experimental groups received MWCNT at doses of 40 and 160 $\mu\text{g}/\text{head}$ (equivalent to 160 or 640 $\mu\text{g}/\text{kg}$ body weight) by intratracheal instillation. The dose levels of MWCNT were selected, in consideration of doses which would allow extrapolation to potential exposure of humans to MWCNT in real workplaces. The positive control group was intratracheally instilled with α -quartz at a dose of 160 $\mu\text{g}/\text{head}$, while the negative control group received PBS-Tween 80 by intratracheal instillation. MWCNT- and vehicle-dosed animals were sacrificed on Day 1, 7, 28 or 91 following intratracheal instillation, whereas the α -quartz-dosed animals were not sacrificed on Day 7. Mean and SD of body weights were 262.2 ± 11.7 g for all the rats immediately before intratracheal instillation.

Lung fixation and histopathology

In order to prepare for light microscopic examination, 8 rats/group were sacrificed at each time point by exsanguination from the jugular vein under pentobarbital anesthesia, and the trachea was ligated. The organs and tissues were fixed by perfusion with physiological saline and subsequently 10% neutral buffered formalin, and embedded in paraffin. Slices of the left lung, 5 μm thick, were cut along the longitudinal axis of the main bronchus. The slices were stained with hematoxylin and eosin (H & E) or Masson's trichrome.

Deposition of MWCNT was semi-quantitatively evaluated for the extent to which the instilled MWCNT was deposited in the bronchiolar space, alveolar space, alveolar wall and bronchus-associated lymphoid tissue (BALT). The severity grade of histopathological changes was scored semi-quantitatively for infiltration of inflammatory cells primarily composed of neutrophils, hyperplasia of Type II pneumocytes, microgranuloma and fibrosis. The extent of MWCNT deposition or the severity grade of histopathological change was scored,

according to the following criteria by microscopic observation of H & E- or Masson's trichrome-stained lung tissues. Score 1, termed "slight", indicates that slight MWCNT deposition or histopathological change was observed in a limited part of the area. Score 2, termed "moderate", indicates that slight MWCNT deposition or histopathological change was observed in a large part of the area or that moderate MWCNT deposition or histopathological change was observed in a limited part of the area. Score 3, termed "marked", indicates that moderate MWCNT deposition or histopathological change was observed in a large part of the area or that marked MWCNT deposition or histopathological change was observed in a limited part of the area. These evaluations were performed by a panel of 3 pathologists certified by the Japanese Society of Toxicologic Pathology.

Measurement of lung weight and biochemical and cytological analyses of BAL fluid

The experimental and control groups of 8 rats each were euthanized under pentobarbital anesthesia. First, the right bronchus was tied, then the right lung was taken out and weighed after lavaging the left lung. The left lung was lavaged 3 times with 7 ml of physiological saline solution, and the wash-out was collected. After the BAL fluid was centrifuged at 10,000 rpm and 4°C for 10 min, aliquots of the acellular supernatant were used for biochemical analysis. Total proteins (TP), albumin, lactate dehydrogenase (LDH) and alkaline phosphatase (ALP) were measured by conventional biochemical methods. TP was chosen as an indicator for maintenance of alveolo-capillary permeability, and LDH and ALP were used as indicators for membrane integrity of pneumocytes and Type II epithelial cells, respectively. The BAL fluid was centrifuged with a cytospin (Cytospin4, Thermo Scientific, USA), and the cellular elements were stained with May-Grünwald-Giemsa. The numbers of mononucleated and multinucleated alveolar macrophages were counted for a total of 1,000 cells under a light-microscope. The presence or absence of MWCNT in the macrophages was also examined.

Statistics

Absolute lung weight and biochemical parameters in the BAL fluid were analyzed by Student's *t*-test, and the number of multinucleated alveolar macrophages in the BAL fluid was analyzed by Dunnett's multiple comparison test, using statistics software (SPSS Japan Inc., Tokyo, Japan). Differences between groups at $p < 0.05$ were considered significant.

Results

Clinical signs and body and lung weights

Neither death nor overt clinical signs were observed in any MWCNT-, α -quartz-dosed or control animals. As shown in Fig. 1, body weights of rats given 40 or 160 μ g MWCNT or 160 μ g α -quartz were not significantly different from those of the two control groups at any time point after intratracheal instillation. The absolute lung weights of the two MWCNT-dosed groups were significantly greater than those of the vehicle control on days 1, 7 and 91 after instillation (Fig. 2). On the other hand, there were no statistically significant differences between the absolute lung weights of the α -quartz-dosed group and the vehicle control (Fig. 2).

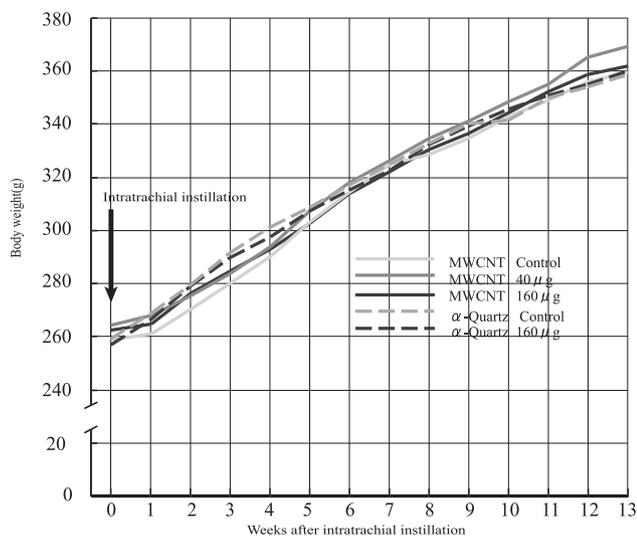


Fig. 1. Temporal changes in body weight of rats which received intratracheal instillation of MWCNT at a dose of 40 or 160 μ g, or α -quartz at a dose of 160 μ g or the vehicle PBS-Tween 80 as positive and negative controls.

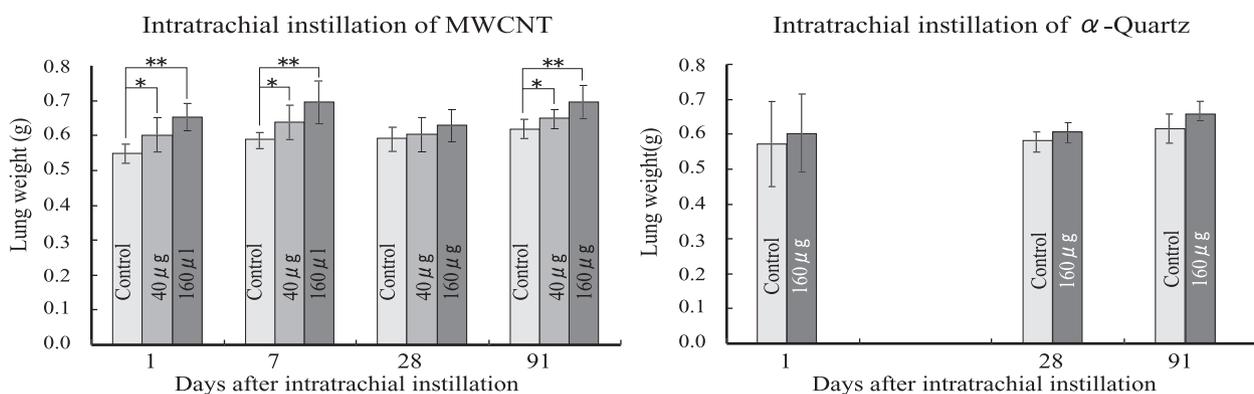


Fig. 2. Absolute lung weights of the rats which received intratracheal instillation of MWCNT at a dose of 40 or 160 μ g/head, or α -quartz at a dose of 160 μ g/head or the vehicle solution as positive and negative controls. Each bar with T indicates mean and S.D. of 8 rats. *: $p < 0.05$ and **: $p < 0.01$ by Student's *t*-test.

Pulmonary deposition of MWCNT

Table 1 shows the pulmonary deposition of MWCNT, and the extent to which MWCNT was deposited in the lung tissues, on different days after instillation. MWCNT deposition was abundant in the bronchiolar and alveolar spaces. The extent of MWCNT deposition decreased in the order of the alveolar space, alveolar wall and BAL. The free form of MWCNT decreased more rapidly in the bronchiolar and alveolar spaces than the phagocytosed form, whereas the phagocytosed form of MWCNT was observed throughout the 91-d post-exposure period. Although MWCNT deposition in the alveolar wall was not observed on Day 1, the deposition of MWCNT and its extent increased on Day 28 and slightly decreased on Day 91. Notably, BAL exhibited a tendency of MWCNT deposition and its extent gradually increased with time after instillation, although MWCNT deposition was not observed on Day 1 in either BAL or the alveolar wall. The extent to which MWCNT was deposited in these lung tissues was milder in the 40- μ g-dosed group than in the 160- μ g-dosed one.

Lung histopathology

Table 2 summarizes the time- and dose-dependent changes and severity of pulmonary toxic responses in the rats given MWCNT by intratracheal instillation at doses of 40 and 160 μ g, with reference to those seen in the 160- μ g α -quartz group, the positive control, and the vehicle PBS group, the negative control. Dose-dependent infiltration of macrophages in the alveolar space persisted throughout the 91-d post-exposure period, whereas infiltration of inflammatory cells composed primarily of neutrophils was found to occur only on Day 1 after dosing 160 μ g MWCNT. Alveolar macrophages without nucleus or with pycnotic nuclei were often found in the bronchiolar space and alveolar

Table 1. Temporal changes in pulmonary deposition of intratracheally instilled MWCNT at a dose of 40 or 160 µg in the rats sacrificed on different days after the instillation

Dose of MWCNT(/rat)	Days after instillation	40 µg				160 µg			
		1	7	28	91	1	7	28	91
No. of rats examined		8	8	8	8	8	8	8	8
Bronchiolar space Non-phagocytosed	Total ^a	8	4	2	0	8	8	2	1
	(Slight Moderate)	(8 0)	(4 0)	(2 0)	(0 0)	(7 1)	(8 0)	(2 0)	(1 0)
Phagocytosed by alveolar macrophages	Total ^a	8	6	7	4	8	8	8	6
	(Slight Moderate)	(8 0)	(6 0)	(7 0)	(4 0)	(7 1)	(8 0)	(8 0)	(6 0)
Alveolar space Non-phagocytosed	Total ^a	8	0	0	0	8	6	0	0
	(Slight Moderate)	(8 0)	(0 0)	(0 0)	(0 0)	(0 8)	(6 0)	(0 0)	(0 0)
Phagocytosed by alveolar macrophages	Total ^a	8	8	8	8	8	8	8	8
	(Slight Moderate Marked)	(0 8 0)	(3 5 0)	(0 8 0)	(7 1 0)	(0 0 8)	(0 0 8)	(0 1 7)	(0 0 8)
	(Slight Moderate Marked)	(0 8 0)	(3 5 0)	(0 8 0)	(7 1 0)	(0 0 8)	(0 0 8)	(0 1 7)	(0 0 8)
Alveolar wall	Total ^a	0	1	8	4	0	8	8	8
	(Slight Moderate)	(0 0)	(1 0)	(8 0)	(3 1)	(0 0)	(8 8)	(8 7)	(8 6)
Bronchus-associated lymphoid tissue	Total ^a	0	3	3	8	0	1	6	8
	(Slight Moderate)	(0 0)	(3 0)	(3 0)	(6 2)	(0 0)	(1 0)	(6 0)	(8 4)

^a:Total number of animals bearing the lesions.
Scoring of grade of pulmonary deposition of MWCNT was given in Materials and Methods.

Table 2. Pulmonary lesions induced by intratracheal instillation of MWCNT at a dose of 40 or 160 µg or α-quartz at a dose of 160 µg in rats sacrificed on different days after the instillation

Dose(/rat)	Days after instillation	MWCNT								α-Quartz				Vehicle control				
		40 µg				160 µg				160 µg								
		1	7	28	91	1	7	28	91	1	7	28	91	1	7	28	91	
No. of rats examined		8	8	8	8	8	8	8	8	8	n.e.	8	8	8	8	8	8	8
Infiltration of alveolar macrophages	Total ^a	8	8	8	8	8	8	8	8	8	-	6	4	0	0	0	0	
	Slight	(0	3	0	7)	(0	0	0	0)	(8	6	4)	(0	0	0	0)		
	Moderate	(8	5	8	1)	(0	0	1	0)	(0	0	0)	(0	0	0	0)		
Infiltration of inflammatory cells	Total ^a	0	0	0	0	8	0	0	0	6	-	0	0	0	0	0	0	
	Slight	(0	0	0	0)	(0	0	0	0)	(6	0	0)	(0	0	0	0)		
	Moderate	(0	0	0	0)	(8	0	0	0)	(0	0	0)	(0	0	0	0)		
Hyperplasia of Type II pneumocytes	Total ^a	0	1	7	4	0	8	8	8	0	-	0	3	0	0	0	0	
	Slight	(0	1	6	4)	(0	8	2	5)	(0	0	3)	(0	0	0	0)		
	Moderate	(0	0	1	0)	(0	0	6	3)	(0	0	0)	(0	0	0	0)		
Microgranuloma	Total ^a	0	0	0	0	0	7	6	7	0	-	0	0	0	0	0	0	
	Slight	(0	0	0	0)	(0	6	1	2)	(0	0	0)	(0	0	0	0)		
	Moderate	(0	0	0	0)	(0	1	5	5)	(0	0	0)	(0	0	0	0)		
Fibrosis	Total ^a	0	0	0	7	0	0	8	8	0	-	1	7	0	0	0	0	
	Slight	(0	0	0	7)	(0	0	8	0)	(0	1	7)	(0	0	0	0)		
	Moderate	(0	0	0	0)	(0	0	0	8)	(0	0	0)	(0	0	0	0)		

Data of vehicle control group are shown for the MWCNT-dosed groups. The vehicle control data for the α-quartz-dosed groups are not shown, since the control group was the same as the vehicle control group of the MWCNT dosed group. Scoring of severity grade of the pulmonary lesions is given in Materials and Methods. n.e.: Not examined. ^a:Total number of animals bearing the lesions.

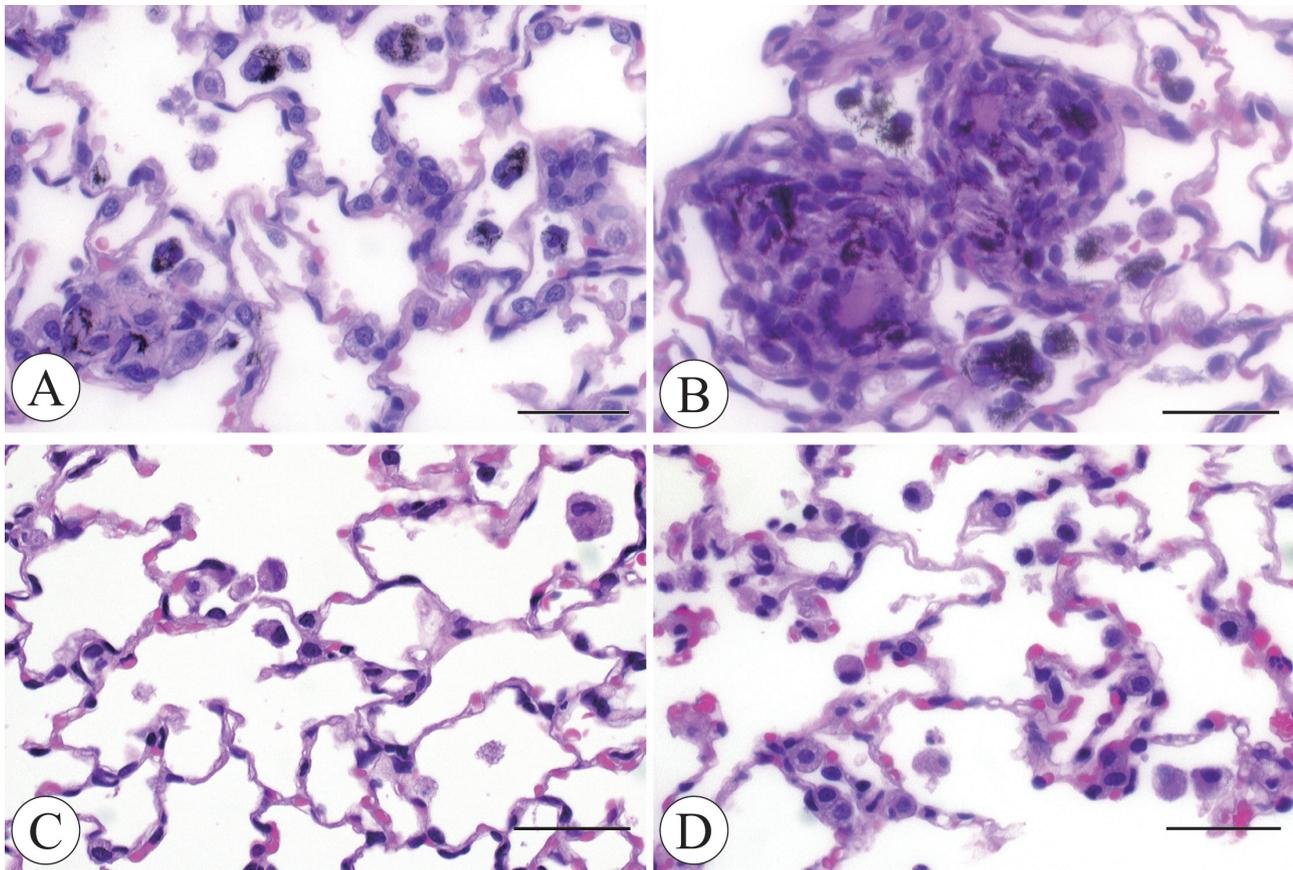


Fig. 3. (A) Hyperplasia of Type II pneumocytes in the alveolar wall of a rat given MWCNT at a dose of 160 μg and sacrificed on Day 28 after instillation. Several alveolar macrophages engulfing MWCNT are seen in the alveolar space. (B) Microgranulomas in the alveolar wall of a rat given MWCNT at a dose of 160 μg and sacrificed on Day 91. (C) Slight infiltration of alveolar macrophages in a rat given α -quartz at a dose of 160 μg and sacrificed on Day 28. (D) Slight infiltration of alveolar macrophages and hyperplasia of Type II pneumocytes in a rat given α -quartz at a dose of 160 μg and sacrificed on Day 91.

H & E stain. Bar indicates 50 μm .

space. Dose-dependent hyperplasia of Type II pneumocytes (Fig. 3A) was observed persistently throughout the 91-d post-exposure period, although hyperplasia did not occur on Day 1. Notably, microgranulomas with diameters up to 150 μm , which were primarily composed of alveolar macrophages engulfing MWCNT (Fig. 3B), were found to occur multifocally only in the 160 μg -dosed group. Some of those macrophages were multinucleated. Although microgranulomas were not seen on Day 1, the number of rats bearing microgranulomas and their severities as expressed by the number lesions in a designated area tended to increase along the time-course of the 91-d post-exposure period. The microgranulomas found in the 160 μg -dosed group were occasionally associated with fibrosis of moderate grade of severity, as evidenced by alveolar wall thickening due to collagen deposition which was stained blue with Masson's trichrome (Fig. 4A). However, slight fibrosis in the absence of overt microgranulomas was

found on Day 91 after instillation of 40 μg MWCNT (Fig. 4B). The pulmonary toxic responses to α -quartz at a dose of 160 μg were characterized by persistent infiltration of macrophages (Fig. 3C and 3D), transient infiltration of inflammatory cells primarily composed of neutrophils, Type II cell hyperplasia (Fig. 3D) and alveolar wall thickening diagnosed as slight fibrosis, all of which were less severe in their severities than the pulmonary toxic responses to MWCNT on an equal mass basis (Table 2). Alveolar wall thickening as slight fibrosis observed on Day 91 after dosing 160 μg α -quartz (Fig. 4C) was similar in grade of severity to that induced on Day 91 after dosing 40 μg MWCNT. No histopathological changes were observed in either bronchi or bronchioles in any treated group. No overt histopathological changes in MWCNT-induced lesions were found in the visceral pleura.

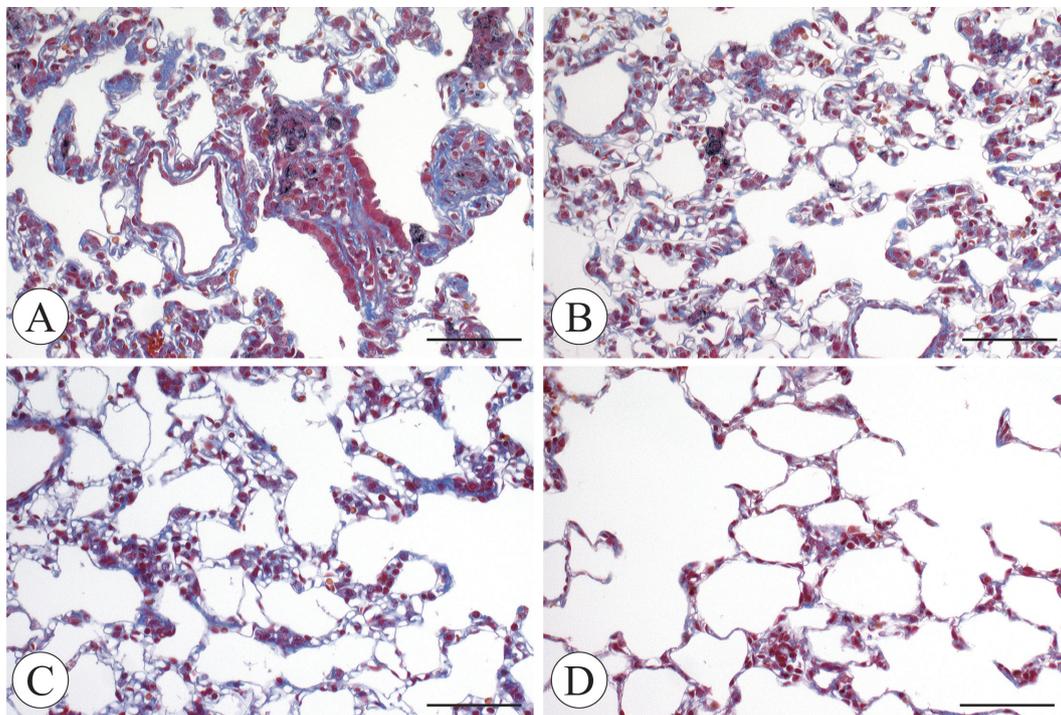


Fig. 4. Fibrosis in the alveolar wall of the rats given MWCNT at a dose of 160 μg (A), 40 μg (B) or α -quartz at a dose of 160 μg (C), and sacrificed on Day 91. A control rat given the vehicle solution at an equal volume and sacrificed on Day 91 (D).

Masson's trichrome stain. Bar indicates 100 μm .

Biochemical and cytological analyses of BAL fluid

TP and albumin contents markedly increased with dose of MWCNT on Day 1. This trend attenuated toward control levels thereafter but persisted significantly until Day 91 (Fig. 5). Temporal changes in both LDH and ALP activities took the same trend as those of TP and albumin. Intratracheal instillation of 160 μg α -quartz induced a significant increase in the contents of proteins and albumin, and LDH and ALP activities on Days 1, 28 and 91 after instillation. Although the biochemical responses to α -quartz on Day 1 were less severe than those to MWCNT, the response magnitudes of α -quartz on Day 91 were similar to those of MWCNT at an equal dose of 160 μg .

Light microscopic examination revealed that a small fraction of alveolar macrophages in the BAL fluid were multinucleated, engulfing the entangled MWCNT fibers in the cytoplasm (inset of Fig. 6). The number of multinucleated alveolar macrophages in the BAL fluid of MWCNT-dosed rats increased significantly and dose-dependently on Day 7 through 91, except on Day 1 (Fig. 6).

Discussion

In the present study, intratracheal instillation of

MWCNT suspended in the PBS-Tween 80 at doses of 40 and 160 $\mu\text{g}/\text{rat}$ was found to produce dose- and time-dependent changes in incidences and severities of pulmonary lesions, lung weight and biochemical and cytological parameters of BAL fluid.

In the separate paper²²⁾, we reported that the 20-min and additional 30-s ultrasonication and intratracheal instillation of MWCNT suspended in PBS containing 0.1% Tween 80 allowed good dispersion of MWCNT fibers in the suspension at the time of intratracheal instillation with a microspray cannula. Besides, the well-dispersed MWCNT fibers were partly engulfed by alveolar macrophages in the alveolar area of MWCNT-dosed rats sacrificed on Day 1 after instillation, suggesting the occurrence of frustrated phagocytosis or incomplete phagocytosis, as proposed by Poland *et al.*⁷⁾ and Hubbs *et al.*²⁴⁾, respectively. Therefore, the results suggest that the tissue around the alveolar area is exposed to well-dispersed MWCNT fibers in the absence of large MWCNT aggregates. Entangled or densely-packed MWCNT fibers in the alveolar interstitium were observed occasionally on Day 91 after instillation. We consider this type of MWCNT to have been formed in the interstitium through re-agglomeration of the MWCNT fibers which had been released from the necrotic or apoptotic macrophages, since in the present

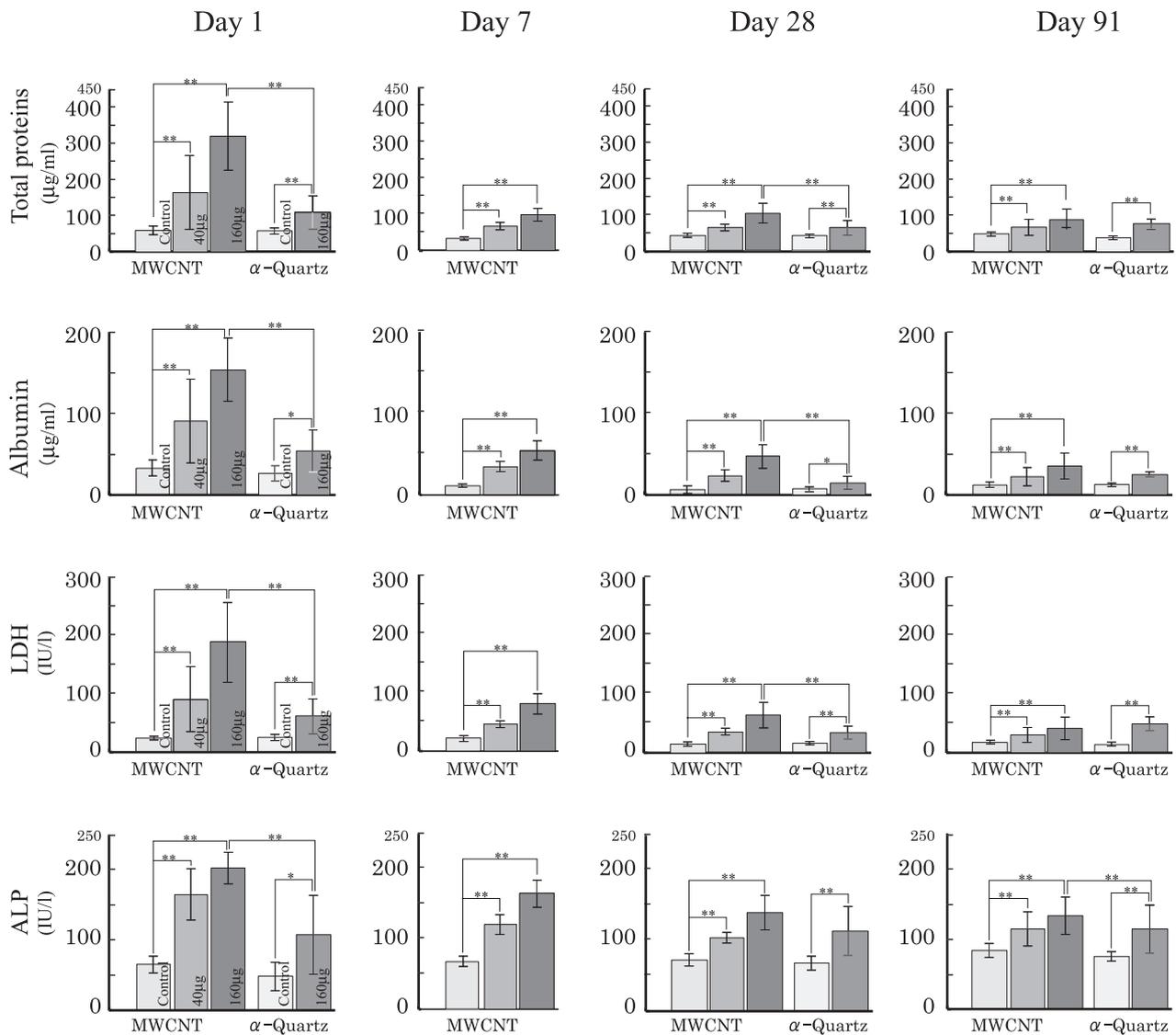


Fig. 5. Contents of total proteins, albumin, LDH and ALP in BAL fluid of the rats which received intratracheal instillation of MWCNT at doses of 40 or 160 µg, or α-quartz at a dose of 160 µg, and were sacrificed on different days after instillation.

A bar with T indicates mean and SD of 8 rats. *: $p < 0.05$ and **: $p < 0.01$ by Student's *t*-test.

study densely-packed MWCNT was not found in the instilling suspension or in the bronchiolar and alveolar spaces on Day 1 after instillation.

Light microscopic observation of pulmonary deposition of MWCNT revealed that the non-phagocytosed (free) form of MWCNT in the bronchiolar and alveolar spaces was cleared faster than the phagocytosed form, and that the phagocytosed form of MWCNT in the bronchiolar and alveolar spaces and the MWCNT deposition in the alveolar wall and BALT, except on Day 1, were persistent throughout the 91-d post-exposure period. Notably, deposition of MWCNT and its extent in the BALT tended to increase along the time course of the 91-d post-exposure period. Morrow²⁵) reported that there are two different pathways of pulmonary lymphatic drain-

age, the pleural drainage and the "deep-set" drainage. BALT belongs to the "deep-set" drainage of periarterial, perivenous and peribronchiolar lymph vessels for clearance of fibers deposited in the interstitium. Pulmonary lymphatic drainage is reported to be much slower in clearing particles and fibers deposited in the interstitium than in the mucociliary escalator²⁶⁻³⁰). Therefore, the present result of a gradual increase in BALT deposition of MWCNT could be accounted by slow clearance of interstitially deposited MWCNT through pulmonary lymphatic drainage unlike the fast clearance of non-phagocytosed MWCNT fibers in the bronchiolar and alveolar spaces through the mucociliary escalator. Persistence of MWCNT fibers in the alveolar interstitium might allow sufficient time for migration of the

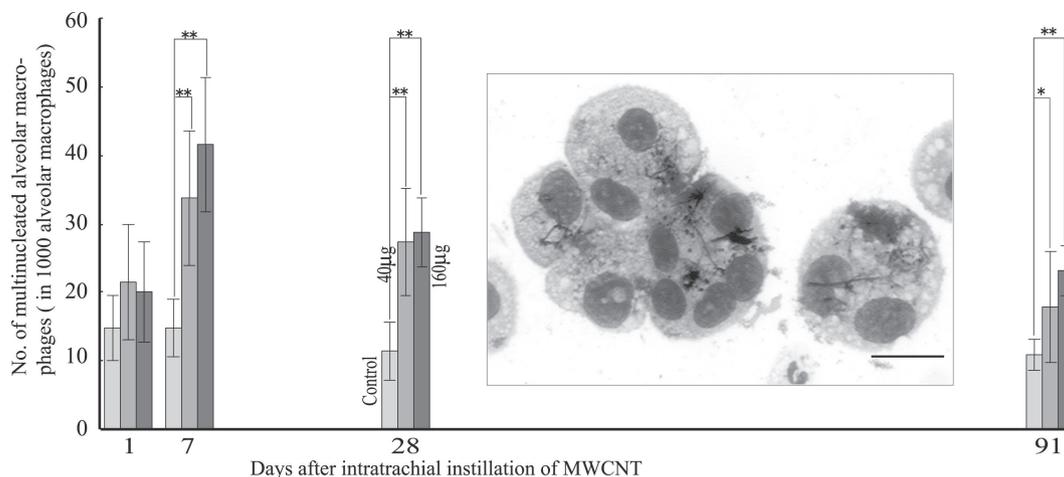


Fig. 6. Number of multinucleated alveolar macrophages in BAL fluid of the rats which received intratracheal instillation of MWCNT at a dose of 40 or 160 μg and were sacrificed on Day 1, 7, 28 or 91 after the instillation. Data indicates means and SDs of 8 rats.

*: $p < 0.05$ and **: $p < 0.01$ by Dunnett's multiple comparison test. Inset shows multinucleated alveolar macrophages engulfing MWCNT in the cytoplasm in the BAL fluid from a rat given 160 μg MWCNT and sacrificed on Day 7 after the instillation. May-Grünwald-Giemsa stain. Bar indicates 25 μm .

fibers through the pulmonary lymphatic pathway adjacent to the pleural cavity. Deposition of MWCNT in the pulmonary lymphatics such as BALT suggests that the dispersed MWCNT fibers might migrate from the alveolar interstitium to the pleura and exert asbestos-like lung pathogenicity in the pleural cavity, as hypothesized by Poland *et al.*⁷⁾ The mean length and width of MWCNT fibers used in the present study were 5.0 μm and 88 nm, respectively, with an averaged aspect ratio of 57, and fibers longer than 5 μm were found to occupy 38.9% of total fibers²²⁾, suggesting a similarity in size and shape of MWCNT and asbestos fibers. Peritoneal and intrascrotal administrations of MWCNT fibers having an equal length and width as those used in the present study are reported to have induced peritoneal mesotheliomas in male p53 gene-deficient mice⁵⁾ and intact male F344 rats⁶⁾, respectively. Intratracheally instilled fibers of chrysotile asbestos are reported to translocate to the pleural cavity in rats³¹⁾, which corresponds to the human parietal pleura as a preferential target of asbestos-related pathology³²⁾.

In the present study, intratracheal instillation of MWCNT at a high dose of 160 μg was found to multifocally induce microgranulomas with diameters up to 150 μm , which were primarily composed of alveolar macrophages engulfing dispersed MWCNT fibers. Besides, the microgranuloma was occasionally associated with fibrosis which was evidenced by alveolar wall thickening due to collagen deposition stained blue with Masson's trichrome. A granulomatous response in close association with inflammation has been addressed

in rodents receiving an agglomerated, less dispersed form of SWCNT or MWCNT at high doses by intratracheal instillation or pharyngeal aspiration. Lam *et al.*⁸⁾ observed that a single intratracheal instillation of SWCNT in mice induced persistent epithelioid granulomas and interstitial inflammation. However, Warheit *et al.*¹⁵⁾ reported that intratracheal instillation of SWCNT in rats given a bolus dose of agglomerated SWCNT induced multifocal granulomas centered around SWCNT, but they ascribed the granulomatous response as non-specific outcome. It was demonstrated that pharyngeal aspiration of agglomerated, less dispersed SWCNT in mice induced granulomatous lesions associated with dense SWCNT aggregate as well as inflammation and diffuse interstitial fibrosis with alveolar wall thickening⁹⁾. However, well-dispersed SWCNT did not produce granulomatous lesions but caused more extensive interstitial fibrosis than less-dispersed SWCNT fibers¹⁰⁾. Muller *et al.*¹²⁾ showed biochemical and histological lines of evidence for inflammation and fibrosis in the lungs of rats given MWCNT by intratracheal instillation. It can be inferred, therefore, that the microgranulomas and slight to moderate fibrosis with alveolar wall thickening found in the present study might have been caused by the persistence of MWCNT in the alveolar wall and interstitium, resulting in persistent infiltration of macrophages which would release proinflammatory and profibrogenic cytokines during phagocytosis.

Hyperplasia of Type II pneumocytes found in the present study is considered to reflect either increased number of Type II cells due to Type I cell injury or

proliferation of Type II cells, or both. This histopathological finding is compatible with the significantly increased ALP activity in the BAL fluid throughout the 91-d post-exposure period, since enhanced ALP activity has been reported to reflect secretory activity of Type II pneumocytes^{15, 33}). The present result is also consistent with the findings of Shvedova *et al.* that pharyngeal aspiration of SWCNT in mice increased the immunofluorescence of cytokeratins 8/18 and markers of Type II epithelial cells in the alveolus, and the number of cells expressing cytoplasmic lamellar bodies in TEM sections. Hyperplasia of Type II pneumocytes and persistent infiltration of macrophages throughout the 91-d post-exposure period, together with the significant increases in total proteins and albumin in the BAL fluid until Day 91, might actually reflect impaired integrity of the alveolar space-capillary function such as gas exchange, resulting in increased alveolo-capillary permeability of proteins and Type II cell hyperplasia, probably due to injury of Type I pneumocytes by MWCNT.

In the present study, the number of multinucleated macrophages engulfing MWCNT in the BAL fluid increased significantly and dose-dependently on Days 7 and 91. This finding is consistent with previously published results that multinucleated macrophages were observed after intraperitoneal injection of MWCNT in mice⁵⁻⁷) and intratracheal instillation of SWCNT in rats¹⁵). The formation of multinucleated alveolar macrophages engulfing MWCNT fibers can be hypothesized to result from interference with the mitotic spindle or steric blocking of cytokinesis by MWCNT fibers, or both. This hypothesis seems to be supported by two previously reported findings with SWCNT^{11, 13}). Mangum *et al.*¹³) reported that a small fraction of alveolar macrophages in the BAL fluid of SWCNT-exposed rats was bridged by intercellular carbon structures that extended into the cytoplasm of each macrophage. Shvedova *et al.*¹¹) suggested possible interference with the mitotic spindle on the basis of their histopathological observation of an anaphase bridge in a dividing macrophage containing SWCNT in the lung of mice exposed by inhalation to SWCNT aerosol. Further evidence in support of this was provided by the formation of polyploidy by sterical blocking of cytokinesis in cultured LLC-MK₂ cells exposed *in vitro* to long crocidolite asbestos fibers³⁴). Since the MWCNT fibers used in the present study were long and thin²²), like asbestos fibers, well-dispersed in the lung tissues and biopersistent, MWCNT might act upon the lung like asbestos fibers, as suggested by Poland *et al.*⁷).

The pulmonary lesions induced by 160 μg α -quartz were characterized by slight but statistically significant increases in the biochemical parameters of the BAL

fluid on Day 1 as compared with the marked lesions induced by 160 μg MWCNT, and by persistent infiltration of macrophages, and slight grade of Type II cell hyperplasia and fibrosis with alveolar wall thickening on Day 91. It has been reported that nodular changes in the interstitium, lipoproteinosis and fibrosis are induced by intratracheal instillation of α -quartz to rats at higher dose levels than 1 mg^{15, 35}) or by repeated inhalation exposure of rats to α -quartz at 15 mg/m³ for 60 d³⁶). However, no such lesions except fibrosis with alveolar wall thickening were found to occur in the lung of 160 μg quartz-dosed rats. Therefore, the comparison between MWCNT and α -quartz findings suggests that MWCNT-induced lesions are more potent on Day 91 than the α -quartz-induced ones on an equal mass basis, although the biochemical responses of BAL fluid to MWCNT were similar in magnitude on Day 91 than those to α -quartz.

Animal data of exposure concentration-response relationships from inhalation exposure are more relevant to the health risk assessment of workers exposed to MWCNT than those from intratracheal instillation, because inhalation exposure is a primary route of exposure for humans handling MWCNT in workplaces. Besides, intratracheal instillation of MWCNT is non-physiological, and involves invasive delivery which bypasses the upper respiratory tract, usually at a dose and/or dose rate substantially greater than that which would have occurred in inhalation. An excessive lung burden due to insoluble and low-toxic particles termed "lung overload" has been reported to impair alveolar macrophage clearance which in turn leads to increased retention and accumulation of particles in the lung, resulting in development of chronic inflammation and fibrosis^{37, 38}). The dose levels that cause the lung overload in rats vary among the related reports. Morrow³⁷) reported that the level of dust burden causing overloading appears to be greater than 1–2 mg of persistently retained dust in the lungs of F344 rats, whereas Driscoll *et al.*³⁹) recommended that intratracheal doses below approximately 100 $\mu\text{g}/\text{rat}$ be used to minimize the interference of clumping and localized inflammatory responses to insoluble particles. Drew *et al.*⁴⁰) reported that single boluses of glass fibers at high doses (2 and 20 mg/rat) produced artifactual granulomatous lesions in the rat lung, whereas repeated exposure to low doses of fiber (0.1 mg/rat) resulted in a fiber distribution and response similar to that after inhalation. Taking these reports³⁷⁻⁴⁰) into consideration, a single dose of 160 $\mu\text{g}/\text{rat}$ for intratracheal instillation of MWCNT, as used in the present study, would minimize the lung overload and induce unambiguous pulmonary toxic responses to MWCNT. In particular, the low dose of 40 $\mu\text{g}/\text{rat}$

would not cause the lung overload.

In the present study, a single intratracheal instillation of well-dispersed MWCNT at a dose of 40 $\mu\text{g}/\text{rat}$ was found to induce slight but persistent changes in the biochemical parameters in BAL fluid and histopathological lung lesions including Type II cell hyperplasia and slight fibrosis with alveolar wall thickening in the absence of overt microgranulomas. Assuming respiratory rate of 561 ml/min/kg body weight⁴¹⁾ for a rat weighing 0.25 kg and a lung deposition efficiency of 11%⁴²⁾ for MWCNT in the alveolar area, the intratracheally instilled dose of 40 $\mu\text{g}/\text{rat}$ can be estimated to be equivalent to the lung burden resulting from inhalation exposure of rats to MWCNT aerosol at 5.4 mg/m³ for 8 h. This estimated airborne level was found to coincide well with the current OSHA's permissible exposure limit (PEL) of 5 mg/m³ TWA (respirable fraction) for graphite particles⁴³⁾.

No data for CNT concentrations in workplace air have been reported to date, except for the measurement⁴⁴⁾ of the peak airborne concentration of respirable dust during handling of SWCNT in a real workplace, 53 $\mu\text{g}/\text{m}^3$ by Maynard *et al.* Assuming that a worker would breathe air laden with MWCNT aerosol at an average of 53 $\mu\text{g}/\text{m}^3$ during an 8-h shift of light work with a minute ventilation of 0.02 m³/min⁴⁵⁾, and assuming that 11% of respirable MWCNT aerosol with an aerodynamic diameter of 0.70 μm would be deposited in the alveolar area⁴⁶⁾, 56 μg of MWCNT would be deposited on the worker's alveolar epithelial surface area. Assuming even distribution of 56 μg of inhaled MWCNT and 40 μg of intratracheally instilled MWCNT over the alveolar surface area of human and rat lungs, respectively, and normalizing to the equivalent alveolar surface area in the human (143 m²/lung) and rat (0.392 m²/lung) from the published morphometric analysis⁴⁷⁾, the amount of MWCNT deposited on the unit alveolar surface area would be 0.39 $\mu\text{g}/\text{m}^2$ for humans and 102 $\mu\text{g}/\text{m}^2$ for rats. With additional assumptions of both no clearance of MWCNT depositing on the surface of alveoli and equal susceptibility of the alveolar epithelium per unit area to MWCNT for humans and rats, the burden of MWCNT on the unit alveolar surface area of the 40 μg MWCNT-instilled rat can be estimated to be equivalent to the alveolar burden of MWCNT of a worker who would breathe air laden with MWCNT aerosol at an average concentration of 53 $\mu\text{g}/\text{m}^3$ for 8 h/day and 260 workdays. Therefore, it is considered that the low dose level of MWCNT used in the present study is relevant to potential occupational exposure to MWCNT, suggesting a need for the establishment of an occupational standard for airborne CNT aerosol in workplace air.

In conclusion, single intratracheal instillations of well-dispersed MWCNT at doses of 40 and 160 $\mu\text{g}/\text{rat}$ were found to induce histopathological, cytological and biochemical changes in the lung tissues and BAL fluid. These MWCNT-induced pulmonary lesions were time- and dose-dependent and more potent than those induced by α -quartz on an equal mass basis. The present results with intratracheally instilled MWCNT were discussed with regard to extrapolation to potential inhalation exposure of humans to MWCNT at workplaces based on several assumptions.

Acknowledgements

The authors are deeply indebted to Dr. Haruhiko Sakurai, Technical Advisor, Occupational Health Research and Development Center, Japan Industrial Safety and Health Association for his fruitful discussion on risk assessment of MWCNT. We also thank Ms. Hitomi Kondo, Misae Saito, Mrs. Hirokazu Kano, Masaaki Suzuki, Dr. Michiharu Matsumoto, Dr. Tetsuya Takeuchi and Dr. Hirokazu Okuda for their excellent technical assistance with the BAL analyses, intratracheal instillation, statistical analysis, and animal care and clinical observation. The present study was supported in part by a Grant-in-Aid for Scientific Research from the Ministry of Health, Labour and Welfare of Japan.

References

- 1) Martin CR, Kohli P (2003) The emerging field of nanotube biotechnology. *Nat Rev Drug Discov* **2**, 29–37.
- 2) Ajayan PM, Charlier JC, Rinzler AG (1999) Carbon nanotubes: from macromolecules to nanotechnology. *Proc Natl Acad Sci USA* **96**, 14199–200.
- 3) Ball P (2001) Roll up for the revolution. *Nature* **414**, 142–4.
- 4) Toray Corporate Business Research, INC. Report on the survey of production and use of nanomaterials in Japan, -Fy 2007-. <http://www.mhlw.go.jp/shingi/2008/04/dl/s0404-3c.pdf>. Accessed July 21, 2009 (in Japanese).
- 5) Takagi A, Hirose A, Nishimura T, Fukumori N, Ogata A, Ohashi N, Kitajima S, Kanno J (2008) Induction of mesothelioma in p53+/- mouse by intraperitoneal application of multi-wall carbon nanotube. *J Toxicol Sci* **33**, 105–16.
- 6) Sakamoto Y, Nakae D, Fukumori N, Tayama K, Maekawa A, Imai K, Hirose A, Nishimura T, Ohashi N, Ogata A (2009) Induction of mesothelioma by a single intrascrotal administration of multi-wall carbon nanotube in intact male Fischer 344 rats. *J Toxicol Sci* **34**, 65–76.
- 7) Poland CA, Duffin R, Kinloch I, Maynard A, Wallace

- WAH, Seaton A, Stone V, Brown S, MacNee W, Donaldson K (2008) Carbon nanotubes introduced into the abdominal cavity of mice show asbestos-like pathogenicity in a pilot study. *Nature Nanotechnology* **3**, 423–8.
- 8) Lam CW, James JT, McCluskey R, Hunter RL (2004) Pulmonary toxicity of single-wall carbon nanotubes in mice 7 and 90 days after intratracheal instillation. *Toxicol Sci* **77**, 126–34.
 - 9) Shvedova AA, Kisin ER, Mercer R, Murray AR, Johnson VJ, Potapovich AI, Tyurina YY, Gorelik O, Arepalli S, Schwegler-Berry D, Hubbs AF, Antonini J, Evans DE, Ku BK, Ramsey D, Maynard A, Kagan VE, Castranova V, Baron P (2005) Unusual inflammatory and fibrogenic pulmonary responses to single-walled carbon nanotubes in mice. *Am J Physiol Lung Cell Mol Physiol* **289**, L698–L708.
 - 10) Mercer RR, Scabilloni J, Wang L, Kisin E, Murray AR, Schwegler-Berry D, Shvedova AA, Castranova V (2008) Alteration of deposition pattern and pulmonary response as a result of improved dispersion of aspirated single-walled carbon nanotubes in a mouse model. *Am J Physiol Lung Cell Mol Physiol* **294**, L87–L97.
 - 11) Shvedova AA, Kisin E, Murray AR, Johnson VJ, Gorelik O, Arepalli S, Hubbs AF, Mercer RR, Keohavong P, Sussman N, Jin J, Yin J, Stone S, Chen BT, Deye G, Maynard A, Castranova V, Baron PA, Kagan VE (2008) Inhalation vs. aspiration of single walled carbon nanotubes in C57BL/6 mice: inflammation, fibrosis, oxidative stress, and mutagenesis. *Am J Physiol Lung Cell Mol Physiol* **295**, L552–L65.
 - 12) Muller J, Huaux F, Moreau N, Misson P, Heilier JF, Delos M, Arras M, Fonseca A, Nagy JB, Lison D (2005) Respiratory toxicity of multi-wall carbon nanotubes. *Toxicol Appl Pharmacol* **207**, 221–31.
 - 13) Mangum JB, Turpin EA, Antao-Menezes A, Cesta MF, Bermudez E, Bonner JC (2006) Single-walled carbon nanotube (SWCNT)-induced interstitial fibrosis in the lungs of rats is associated with increased levels of PDGF mRNA and the formation of unique intercellular carbon structures that bridge alveolar macrophages *in Situ*. *Particle Fibre Toxicol* **3**, 1–13.
 - 14) Shvedova AA, Kisin ER, Murray AR, Gorelik O, Arepalli S, Castranova V, Young S-H, Gao F, Tyurina YY, Oury TD, Kagan VE (2007) Vitamine E deficiency enhances pulmonary inflammatory response and oxidative stress induced by single-walled carbon nanotubes in C57BL/6 mice. *Toxicol Appl Pharmacol* **221**, 339–48.
 - 15) Warheit DB, Laurence BR, Reed KL, Roach DH, Reynolds GAM, Webb TR (2004) Comparative pulmonary toxicity assessment of single-wall carbon nanotubes in rats. *Toxicol Sci* **77**, 117–25.
 - 16) Elgrabli D, Abella-Gallart S, Robidel F, Rogerieux F, Boczkowski J, Lacroix G (2008) Induction of apoptosis and absence of inflammation in rat lung after intratracheal instillation of multiwalled carbon nanotubes. *Toxicol* **253**, 131–6.
 - 17) Mitchell LA, Gao J, Wal RV, Gigliotti A, Burchiel SW, McDonald JD (2007) Pulmonary and systemic immune response to inhaled multiwalled carbon nanotubes. *Toxicol Sci* **100**, 203–14.
 - 18) Muller J, Huaux F, Fonseca A, Nagy JB, Moreau N, Delos M, Raymundo-Piñero E, Béguin F, Kirsch-Volders M, Fenoglio I, Fubini B, Lison D (2008) Structural defects play a major role in the acute lung toxicity of multiwall carbon nanotubes: toxicological aspects. *Chem Res Toxicol* **21**, 1698–705.
 - 19) Muller J, Decordier I, Hoet PH, Lombaert N, Thomassen L, Huaux F, Lison D, Kirsch-Volders M (2008) Clastogenic and aneugenic effects of multi-wall carbon nanotubes in epithelial cells. *Carcinogenesis* **29**, 427–33.
 - 20) Hubbard R, Venn A, Lewis S, Britton J (2000) Lung cancer and cryptogenic fibrosing alveolitis. A population-based cohort study. *Am J Respir Crit Care Med* **161**, 5–8.
 - 21) Fraire AE, Greenberg SD (1973) Carcinoma and diffuse interstitial fibrosis of lung. *Cancer* **31**, 1078–86.
 - 22) Takaya M, Serita F, Yamazaki K, Aiso S, Kubota H, Asakura M, Ikawa N, Nagano K, Arito H, Fukushima S (2010) Characteristics of multiwall carbon nanotubes for an intratracheal instillation study with rats. *Ind Health* **48**, 452–9.
 - 23) National Research Council (1996) Guide for the care and use of laboratory animals. National Academy Press, Institute of Laboratory Animal Resources Commission on Life Sciences, NRC, Washington, DC.
 - 24) Hubbs A, Mercer RR, Coad JE, Barrelli LA, Willard PA, Sriram K, Wolfarth M, Castranova V, Porter D (2009) Persistent pulmonary inflammation, airway mucous metaplasia and migration of multiwalled carbon nanotubes from the lung after subchronic exposure. *The Toxicologist* (48th Annual Meeting of SOT) 457.
 - 25) Morrow PE (1972) Lymphatic drainage of the lung in dust clearance. *Ann NY Acad Sci* **200**, 46–65.
 - 26) Lee KP, Trochimowicz HJ, Reinhardt CF (1985) Transmigration of titanium dioxide (TiO₂) particles in rats after inhalation exposure. *Exp Mol Pathol* **42**, 331–43.
 - 27) Takahashi S, Patrick G (1987) Patterns of lymphatic drainage to individual thoracic and cervical lymph nodes in the rat. *Lab Anim* **21**, 31–4.
 - 28) Lehnert BE, Valdez YE, Stewart CC (1986) Translocation of particles to the tracheobronchial lymph nodes after lung deposition: kinetics and particle-cell relationships. *Exp Lung Res* **10**, 245–66.
 - 29) Ferin J, Oberdörster G, Penney DP (1992) Pulmonary retention of ultrafine and fine particles in rats. *Am J Respir Cell Mol Biol* **6**, 535–42.
 - 30) Warheit DB, Hansen JF, Yuen IS, Kelly DP, Snajdr SI, Hartskey MA (1997) Inhalation of high concentrations of low toxicity dusts in rats results in impaired pul-

- monary clearance mechanisms and persistent inflammation. *Toxicol Appl Pharmacol* **145**, 10–22.
- 31) Viallat JR, Raybuad F, Passarel M, Boutin C (1986) Pleural migration of chrysotile fibers after intratracheal injection in rats. *Arch Environ Health* **41**, 282–6.
 - 32) Suzuki Y, Kohyama N (1991) Translocation of inhaled asbestos fibers from the lung to other tissues. *Am J Ind Med* **19**, 701–4.
 - 33) Miller BE, Hook GER (1990) Hypertrophy and hyperplasia of alveolar type II cells in response to silica and other pulmonary toxicants. *Environ Health Perspect* **85**, 15–23.
 - 34) Jensen CG, Jensen LCW, Rieder CL, Cole RW, Ault JG (1996) Long crocidolite asbestos fibers cause polyploidy by sterically blocking cytokinesis. *Carcinogenesis* **17**, 2013–21.
 - 35) Gross KB, White HJ, Smiler KL (1984) Functional and morphologic changes in the lungs after a single intratracheal instillation of silica. *Am Rev Respir Dis* **129**, 833–9.
 - 36) Porter DW, Hubbs A, Mercer R, Robinson VA, Ramsey D, McLaurin J, Khan A, Battelli L, Brumbaugh K, Teass A, Castranova V (2004) Progression of lung inflammation and damage in rats after cessation of silica inhalation. *Toxicol Sci* **79**, 370–80.
 - 37) Morrow PE (1988) Possible mechanisms to explain dust overloading of the lungs. *Fundam Appl Toxicol* **10**, 369–84.
 - 38) Oberdörster G (1995) Lung particle overload: implications for occupational exposures to particles. *Regul Toxicol Pharmacol* **27**, 123–35.
 - 39) Driscoll KE, Costa DL, Hatch G, Henderson R, Oberdörster G, Salem H, Schlesinger B (2000) Intratracheal instillation as an exposure technique for the evaluation of respiratory tract toxicity: uses and limitations. *Toxicol Sci* **55**, 123–35.
 - 40) Drew RT, Kuschner M, Bernstein DM (1987) The chronic effects of exposure of rats to sized glass fibers. *Ann Occup Hyg* **31**, 711–29.
 - 41) Mauderly JL, Tesarek JE, Sifford LJ, Sifford LJ (1979) Respiratory measurements of unsedated small laboratory mammals using nonbreathing valves. *Lab Anim Sci* **29**, 323–9.
 - 42) Leong BK, Sabaitis CP, Rop DA and Aaron CS (1998) Quantitative morphometric analysis of pulmonary deposition of aerosol particles inhaled via intratracheal nebulization, intratracheal instillation or nose-only inhalation in rats. *J Appl Toxicol* **18**, 149–60.
 - 43) Occupational Safety and Health Administration (2006) Regulation (Standards–29 CFR). Part 1910, occupational safety and health standards. Toxic and hazardous substances. Table Z-1 Limits for air contaminants. OSHA, Washington, DC. http://www.osha.gov/pls/oshaweb/owadisp.show_document?p_table=STANDARDS&p_id=9992#. Accessed July 21, 2009.
 - 44) Maynard AD, Baron PA, Foley M, Shvedova AA, Kisin ER, Castranova V (2004) Exposure to carbon nanotube material: aerosol release during the handling of unrefined single-walled carbon nanotube material. *J Toxicol Environ Health, Part A* **67**, 87–107.
 - 45) Galer DM, Leung HW, Sussman RG, Trzos RJ (1992) Scientific and practical considerations for the development of occupational exposure limits (OELs) for chemical substances. *Regul Toxicol Pharmacol* **15**, 291–306.
 - 46) International Commission on Radiological Protection (2002) Reference values for regional deposition. In: *Annals of the ICRP. ICRP supporting guidance 3. Guide for the practical application of the ICRP human respiratory tract model*, Valentin J (Ed.), 91–2, Pergamon (Elsevier), Oxford.
 - 47) Pinkerton KE, Gehr P, Crapo JD (1991) Architecture and cellular composition of the air-blood barrier. In: *Treatise on pulmonary toxicology. Vol. 1. Comparative biology of the normal lung*, Parent RA (Ed.), 121–8. CRC Press, Boca Raton.